

Isolation and *in vitro* Activation of Mouse Peyer's Patch Cells from Small Intestine Tissue

Claudia Pastori and Lucia Lopalco*

Division of Immunology, San Raffaele Scientific Institute, Milan, Italy

*For correspondence: lopalco.lucia@hsr.it

[Abstract] The lumen of gastrointestinal tract is exposed to several potentially pathogenic microorganisms, thus it is extremely relevant to understand how immunosurveillance can be established. Peyer's Patches (PPs) are oval or round lymphoid nodules that protrude from the outer wall of the ileum portion of small intestine. PPs contain a high percentage of B and T lymphocytes, macrophages and dendritic cells. Here we summarize a protocol for isolation and culture of mouse PP cells, which can be used to get a better insight into immunopathologies of microbes and to evaluate immune responses elicited by mucosal vaccines.

Materials and Reagents

1. Mouse (e.g. Balb/c) 8-20 weeks old
2. Sterile PBS without Ca/Mg (Euroclone, catalog number: ECB4004L)
3. Pen/Strep: 100x solution of penicillin/streptomycin/fungizone (Life Technologies, Gibco®, catalog number: 15240-062)
4. Gentamycin (50 mg/ml solution) (Life Technologies, Gibco®, catalog number: 15750-037)
5. RPMI 1640 (with glutamine) (Lonza, catalog number: BE12-702F)
6. Fetal bovine serum (FBS) (heat inactivated) (Lonza, catalog number: DE14-801F)
7. 0.4% trypan blue (Sigma-Aldrich, catalog number: T8154)
8. Phorbol 12-Myristate 13-Acetate (PMA) (Sigma-Aldrich, catalog number: P8139 - follow the manufacturer's instructions to prepare stock solutions)
9. Ionomycin (Sigma-Aldrich, catalog number: I9657 - follow the manufacturer's instructions to prepare stock solutions)
10. Common disinfectant (*i.e.* 70-90% ethanol, tincture of iodine)
11. PBS + 2% Pen/Strep (see Recipes)
12. RPMI + 2% Pen/Strep (see Recipes)
13. Complete medium (see Recipes)

Equipment

1. Soft wood tablet and pins

2. Aluminium foils (or another smooth, non-absorbent surface)
3. Three sterile small thin surgical scissors
Note: In particular one for cutting the skin, the second one for cutting the muscular wall and the intestine, and the last one for removing PPs.
4. Four sterile small thin surgical tweezers, two straight and two curved (straight ones for cutting the skin, the muscular wall and the intestine; curved ones to expel faeces and to remove PPs)
5. 70 µm cell strainer (BD Biosciences, Falcon®, catalog number: 352350)
6. 2.5 ml syringes
7. 15 and 50 ml centrifuge tubes
8. 24-well cell culture plates
9. Refrigerated centrifuge
10. Sterile flow hood
11. Thermostatic water bath
12. Inverted microscope for cell cultures
13. 37 °C, 5% CO₂ cell culture incubator

Procedure

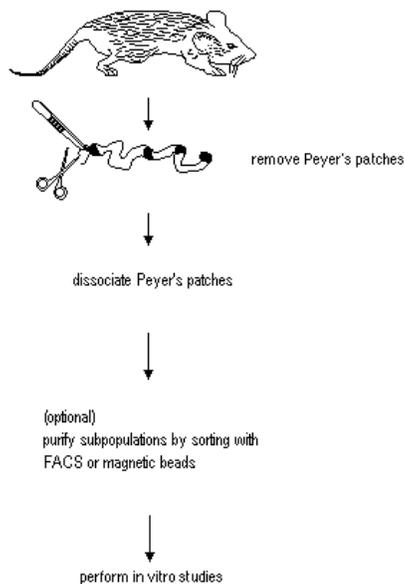


Figure 1. Diagram for the isolation of PP cells from small intestine. Modified from Lefrancois and Lycke (2001).

- A. Removal of small intestine and excision of Peyer's patches
1. Euthanize mouse by CO₂ asphyxiation, following guide lines approved by your Institutional Animal Care and Use Committee. Proceed as soon as possible to the aseptic intestine explantation and PP dissociation to obtain maximum cell viability.
Note: All subsequent steps should be performed in a sterile flow hood.
 2. Gently lay down the mouse on its back, on a soft wood surface, stretch the limbs and fix the four paws with pins.
 3. Clean the abdomen with the disinfectant.
 4. A pair of scissors and a straight tweezers are used to perform a midline incision and retract the skin, then open the muscular wall with another cutting along median axis with a new pair of scissors and tweezers.
 5. Cut small intestine approximately 0.5 cm below the stomach, draw out of peritoneal cavity unfolding it. Cut intestine about 1 cm above the caecum, then remove fat, mesenteric lymph nodes and adjacent tissues.
Note: You can cut the intestine with the same scissors and tweezers used for the muscular wall.
 6. Place the intestine on an aluminium foil and continuously moisten with cold PBS + 2% Pen/Strep to prevent drying.
 7. Expel faeces by placing a curved tweezers flat on the surface of the intestine and press down along the entire length. Press gently to avoid breaking the intestine.
 8. Remove PPs by tightening the patch with a new curved tweezers and cutting it with scissors, avoiding as much as possible the surrounding intestinal wall. The patches appear as small protruding whitish/greyish nodules (diameter 1-2 mm), similar to the lymph nodes, embedded in the outer intestinal wall, which appears to be yellow-brown. In general, five to ten PPs can be obtained from a single mouse, depending of age and strain.
Notes:
 - a. *The presence of an excessive amount of surrounding epithelium reduces the yield of recovered cells, as it could prevent a good mechanical dissociation.*
 - b. *The removal of the surrounding tissue is crucial if the enzymatic dissociation is carried out; in this last case the intraepithelial cells could contaminate the resulting PP-single cells.*
 9. Immediately after excision, save all PPs in 1 ml cold RPMI + 2% Pen/Strep.
- B. Isolation of PP cells by mechanical dissociation
1. Transfer all PPs in a 70 µm cell strainer placed over a 50 ml tube.

2. Dissociate PPs using the plunger of a 2.5 ml syringe and gently crush the patches forcing through the filter. During dissociation wash the filter with 5 ml of cold RPMI + 2% Pen/Strep (medium flow through the filter contains the single cells of PPs).
3. Repeat step 2 twice more, until they are no longer visible pieces of tissue. At this point, the tube contain PP-single cells flowed through the filter and suspended in 15 ml of RPMI + 2% Pen/Strep.
4. Centrifuge at 400 x g, 4 °C for 10 min. Discard the supernatant, resuspend the cells and add 10 ml of cold RPMI + 2% Pen/Strep.
5. Repeat step 4 two more times. After the last centrifugation resuspend the cells in 1-2 ml of cold RPMI + 2% Pen/Strep.
6. Count viable cells using trypan blue dye exclusion.

Notes:

- a. *The variability in the number of cells recovered from a single mouse is very high, mainly due to the mouse (strain and age), the time elapsed between the euthanization and dissociation of PP, and the accuracy in removing every single PP.*
- b. *The mechanical dissociation allows to isolate mainly non-adherent cells, such as B and T lymphocytes. Adherent cells including monocytes, dendritic and non-immune epithelial cells appear to be present in small quantities since they require an enzymatic treatment to be isolated. The phenotype of recovered PP cells can be checked by FACS analysis.*
- c. *If needed, PP cells can be further purified by sorting with magnetic beads or FACS [see Guo et al. (2008) and Thornton (2003) in the reference section].*

C. *In vitro* activation of PP cells

1. Centrifuge an appropriate amount of cells at 400 x g, at 4 °C for 10 min.
2. Resuspend the pellet in complete medium at a concentration of 2 x 10⁶ cells/ml and culture the PP cells in 24-well plate, 1 ml/well, in presence of 10 ng/ml PMA and 500 ng/ml Ionomycin for at least 24 h, in a 37 °C and 5% CO₂ incubator.

Note: Other activation protocols, specific for B or T lymphocytes, can be used [see Li et al. (2006) and Muul et al. (2011) in the references section].

Optionals:

- a. *To eliminate adherent cells, PP cells can be cultured for 2-4 h in Complete Medium (see recipes) with PMA and Ionomycin without antibiotics. The presence of adherent cells (monocytes and/or dendritic cells) may differentially modulate the activation of B and T lymphocytes with a consequent alterations of the immune responses.*
- b. *To avoid as much as possible a bacterial contamination, PP cells are washed and cultured for 24 h as describe above, in Complete Medium + PMA/Ionomycin, with 100*

µg/ml Gentamycin instead of Penicillin/Streptomycin. The use of a different antibiotic ensures the elimination of all kinds of bacteria.

- After 24 h (or more) the supernatants of cultures can be collected, centrifuged at 400 x g, at 4 °C for 10 min to eliminate the cells, and used to test the presence of specific antibodies or cytokines. Activated cells can be harvested, washed by centrifugation at 4 °C for 10 min and used in flow cytometry assays and sorting.

Representative data

FACS analysis of *in vitro* activated PP cells.

PPs were obtained from 6 Balb/c mice as previously published (Pastori *et al.*, 2014). In detail, two different immunization protocols were used: 3 mice received one intranasal (i. n.), while the other 3 mice received one intraperitoneal (i. p.) immunization. Aluminium hydroxide was used as adjuvant in i. p. immunization. For immunization Flock House Virus (FHV) was used in which an external loop of CCR5 (a seven transmembrane protein, belonging to chemokine receptor family and coreceptor for HIV-1) was introduced. The detailed immunization protocol has been described in Pastori *et al.* (2014).

After excision, PPs were mechanically dissociated and PP cells were *in vitro* activated with PMA and Ionomycin, as describe above. After 24 h cells were harvested and stained with antibodies against some markers of B cell activation.

As shown in Figure 2, CD138 and CD40/CD19 expression indicate a B cell activated phenotype for both protocols.

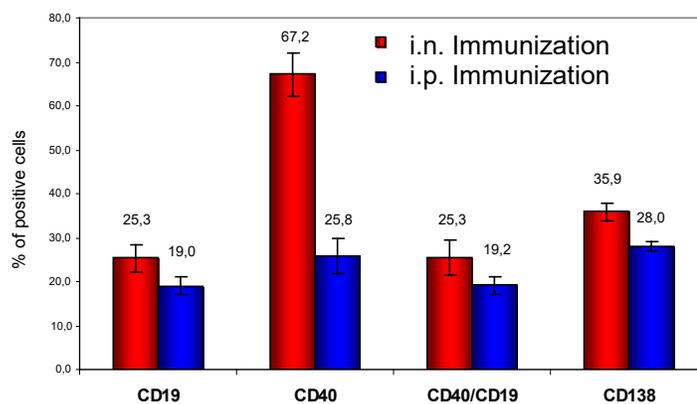


Figure 2. Expression of CD19, CD40, CD138 and co-expression of CD19/CD40 on PP cells after 24 h of PMA/Ionomycin activation. Percentage of positive cells refers to lymphocytes (gated on FSC vs SSC plot). Mean and standard deviation are shown.

Notes

1. Images and more detailed information about mouse necropsy, intestine explantation and Peyer' patches can be found in Scudamore (2013).
2. The yield of cells isolated from PPs or intestinal lymph nodes is time sensitive. Proceed with the isolation as quickly as possible to obtain better cell viability.
3. The mechanical dissociation of PPs gives a lower yield compared to the enzymatic one, but it does not alter the expression of surface antigens, thus, it is better in the case of subsequent FACS analysis or *in vitro* assays. The mechanical dissociation also leads to a relative loss of adherent accessory cells, such monocytes and/or dendritic cells, therefore, for the isolation of these cell types the enzymatic dissociation is preferable.

Recipes

Note: Prepare and keep all solutions in a sterile condition.

1. PBS + 2% Pen/Strep
For 100 ml combine 98 ml of PBS and 2 ml of Pen/Strep
Refrigerate at 4 °C before use
2. RPMI + 2% Pen/Strep
For 100 ml combine 98 ml of RPMI and 2 ml of Pen/Strep
Refrigerate at 4 °C before use
3. Complete medium
For 100 ml combine 89 ml of RPMI (with Glutamine), 10 ml of heat inactivated FBS and 1 ml of Pen/Strep
Warm at 37 °C in a water bath before use

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