

## Measurement of the Intracellular Calcium Concentration with Fura-2 AM Using a Fluorescence Plate Reader

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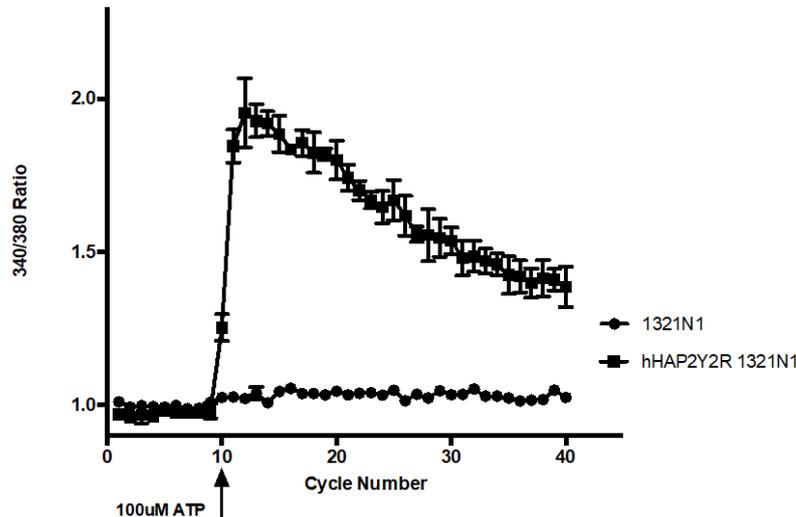
**[Abstract]** Intracellular calcium elevation triggers a wide range of cellular responses. Calcium responses can be affected or modulated by membrane receptors mutations, localization, exposure to agonists/antagonists, among others (Burgos *et al.*, 2007; Martínez *et al.*, 2016). Changes in intracellular calcium concentration can be measured using the calcium sensitive fluorescent ratiometric dye fura-2 AM. This method is a high throughput way to measure agonist mediated calcium responses.

**Keywords:** Cell biology, Cell based analysis, Ion analysis, Calcium, Mammalian cell line, Astrocyte, Physiology

**[Background]** Activation of G protein coupled receptors triggers production in hundreds or thousands of second messenger molecules. Once stimulated,  $G_q$  protein coupled receptors activate phospholipases C (PLC), which cleaves phosphatidylinositol 4,5-bisphosphate ( $PIP_2$ ) into diacyl glycerol (DAG) and inositol 1,4,5-trisphosphate ( $IP_3$ ). DAG remains membrane bound and  $IP_3$  is released into the cytosol where it binds to a specific  $IP_3$  receptor (calcium channel) in the endoplasmic reticulum (ER). This causes an increase of intracellular calcium concentration that modulates the activation of calcium binding proteins and Protein Kinase C (PKC). Therefore, given the importance of calcium in biological systems, many techniques/methods to analyze and measure  $Ca^{2+}$  levels have been established.

To determine the intracellular  $Ca^{2+}$  concentration, fluorescent indicators are particularly useful.  $Ca^{2+}$  indicators are available with variations in affinity, brightness or spectral characteristics. Fura-2-acetoxymethyl ester (fura-2 AM), is a membrane-permeable, non-invasive derivative of the ratiometric calcium indicator fura-2. Fura-2 AM crosses cell membranes and once inside the cell, the cellular esterases remove the acetoxymethyl group.  $Ca^{2+}$ -bound fura-2 AM has its excitation maximum at 335 nm,  $Ca^{2+}$ -free fura-2 AM has its excitation maximum at 363 nm. In both states, the emission maximum is about 510 nm. The typical excitation wavelengths used are 340 nm and 380 nm for  $Ca^{2+}$ -bound and  $Ca^{2+}$ -free fura-2 AM, respectively. The ratios 510 nm/340 nm and 510 nm/380 nm are directly related to the amount of intracellular  $Ca^{2+}$  (Figure 1). Traditionally, changes in intracellular  $Ca^{2+}$  mobilization/concentration were measured using fluorescent calcium indicators and a fluorescent confocal microscope, known as calcium imaging. This protocol explains how to use the Tecan Infinite M200<sup>®</sup> microplate reader equipped with an injector to measure intracellular  $Ca^{2+}$  concentration. With the aid of an injector equipped microplate reader, researchers may be able to screen new agonists for  $G_q$  protein couple receptors and evaluate how the intracellular calcium mobilization is affected by mutant

receptors. This protocol can be adapted to measure agonist mediated intracellular calcium mobilization in several cellular/receptor/agonist models using the appropriate growth conditions.



**Figure 1. The human P2Y2 receptor mobilizes intracellular  $\text{Ca}^{2+}$  efficiently when expressed in 1321N1 human astrocytoma cells.** Representative traces of intracellular calcium responses to 100  $\mu\text{M}$  ATP in 1321N1 cells transfected with hHA-P2Y<sub>2</sub>R (solid squares) and control non-transfected 1321N1 cells (solid circles). Data represent the mean  $\pm$  SEM of readings from 4 independent experiments.

### Materials and Reagents

1. 15 ml centrifuge tube (VWR, catalog number: 89004-368)
2. Clear flat-bottom black 96-well culture trays with lid (Corning, catalog number: 3904)
3. Wild type (WT) human 1321N1 astrocytoma cells devoid of P2 receptors and hHA-P2Y<sub>2</sub>R expressing 1321N1 cells (gift from Dr. Gary A. Weisman, University of Missouri)
4. DPBS
5. Trypsin (TrypLE Express) (Thermo Fisher Scientific, Gibco™, catalog number: 12604013)
6. Probenecid (water soluble) (Thermo Fisher Scientific, Invitrogen™, catalog number: P36400)
7. Adenosine 5'-triphosphate disodium salt (ATP) (Tocris Bioscience, catalog number: 3245)
8. Sodium chloride (NaCl) (Sigma-Aldrich, catalog number: S7653)
9. Potassium chloride (KCl) (Sigma-Aldrich, catalog number: P9333)
10. Calcium chloride ( $\text{CaCl}_2$ ) (Sigma-Aldrich, catalog number: C1016)
11. Magnesium chloride ( $\text{MgCl}_2$ ) (Sigma-Aldrich, catalog number: M8266)
12. HEPES (Sigma-Aldrich, catalog number: H0887)
13. Glucose (Sigma-Aldrich, catalog number: G8270)
14. Bovine serum albumin (BSA) (Sigma-Aldrich, catalog number: A2153)

15. Fura-2-acetoxymethyl ester (fura-2 AM) (Thermo Fisher Scientific, Invitrogen™, catalog number: F1225)
16. Pluronic F-127, 20% (w/v) stock in DMSO (Thermo Fisher Scientific, Invitrogen™, catalog number: P3000MP)
17. Dulbecco's modified Eagle's medium–low glucose (DMEM) (Sigma-Aldrich, catalog number: D2902)
18. Fetal bovine serum (FBS) (Sigma-Aldrich, catalog number: F6178)
19. Antibiotic/antimycotic (Sigma-Aldrich, catalog number: A5955)
20. Geneticin, G418 (Thermo Fisher Scientific, Gibco™, catalog number: 10131035)
21. HEPES buffered saline (HBS), pH 7.4 (see Recipes)
22. Dye solution (see Recipes)
23. Medium for wild type (WT) human 1321N1 astrocytoma cells (see Recipes)
24. Medium for hHA-P2Y<sub>2</sub>R expressing 1321N1 cells (see Recipes)
25. Low serum medium (see Recipes)

## **Equipment**

1. Tecan Infinite M200® (Tecan Trading, model: Infinite® M200)
2. Fisher Scientific Mini vortex (Fisher Scientific)
3. Eppendorf 5810R Benchtop centrifuge (Eppendorf, model: 5810 R)
4. Biorad TC10 cell counter® (Bio-Rad Laboratories, model: TC10™ Automated Cell Counter)  
*Note: This product has been discontinued.*
5. Fisher Scientific Isotemp CO<sub>2</sub> Incubator (5% CO<sub>2</sub>, 37 °C) (Fisher Scientific)
6. Laminar flow biological hood

## **Software**

1. i-Control® (Tecan Trading AG, Switzerland)
2. Excel® (Microsoft)
3. GraphPad Prism® (GraphPad Software Inc., La Jolla, CA)

## **Procedure**

### A. Cell culture and fura-2 loading

1. Maintain cells with the appropriate media in an incubator (37 °C, 5% CO<sub>2</sub>).
2. Grow cell to a 90% confluency, discard the cell medium, wash with DPBS and detach with trypsin.
3. Re-suspend detached cells with 5 ml of cell culture medium and transfer to a 15 ml centrifuge tube.

4. Centrifuge at 300 x g for 5 min.
5. Discard cell culture medium and re-suspend cells with 5 ml of low serum medium.
6. Seed cells 16 h before experiments in clear flat-bottom black 96-well culture trays at a density of  $3.0 \times 10^4$  cells/well (80-90% confluent).
7. Discard the medium by carefully inverting the plate and wash once with 200  $\mu$ l HEPES-buffered saline. Carefully discard HBS by inverting the plate, replace with dye solution and incubate cells 1 h at RT in the dark.
8. Remove the dye solution by inverting the plate, wash twice with HBS and cover with 200  $\mu$ l of HBS supplemented with 2.5 mM probenecid for at least 20 min at RT to allow for fura-2 AM de-esterification.

*Note: Cell medium must be without phenol red to avoid fluorescence measurement interference. To avoid evaporation remember to put plate lid back during incubations.*

#### B. Instrument setup

1. Open the Tecan i-Control software and enter the following parameters:

**Table 1. Dual wavelength Ca<sup>2+</sup> indicator assay measurement parameters and instrument settings for the Tecan Infinite M200<sup>®</sup> with injector.** Optimized settings to measure ATP stimulated intracellular Ca<sup>2+</sup> mobilization in 1321N1 astrocytoma cells.

Measurement parameter	Instrument setting
Plate	[COR96fb clear bottom]–Corning 96 Flat black, clear bottom, with lid
Temperature	28 °C
Wait for temperature	Minimum: 27.5 °C, Maximum: 28.5 °C
Part of plate	Select wells to be measured
Well	
Kinetic cycle	Number of cycles: 40, Kinetic interval: 3 sec
Kinetic condition	Execute commands at cycle 10
Injection	Injector A Volume: 20 $\mu$ l, Speed: 100 $\mu$ l/sec
Fluorescence intensity	Excitation: 380 nm, Emission: 510 nm, Mode: Bottom, Number of flashes: 25, Settle Time: 0 msec, Gain: 120, Integration time: 40 $\mu$ sec
Fluorescence intensity	Excitation: 340 nm, Emission: 510 nm, Mode: Bottom, Number of flashes: 25, Settle Time: 0 msec, Gain: 120, Integration time: 40 $\mu$ sec

2. Before performing the assay, it is recommended to clean the injectors and tubing first with distilled water, then with HBS.
3. Prime injector A with 1.1 mM ATP solution.
4. Set the measurement parameters, insert the plate, remove the plate lid and start the run. Determine fluorescence emission intensity at 510 nm in individual wells alternating excitation wavelengths of 340 and 380 nm every 3 sec for 40 acquisition cycles, using the parameters specified in Table 1.

## **Data analysis**

1. To express in intracellular calcium concentration in cells, calculate the ratio of 510 nm emission, in response to 340/380 nm excitation.
2. Normalize Ca<sup>2+</sup> mobilization using the average of the first 9 cycles (unstimulated) as baseline.
3. See [Appendix](#) for data processing example.

## **Notes**

1. Fura-2 AM, under certain staining conditions can accumulate into cell organelles (endoplasmic reticulum, endosome/lysosome, mitochondrion, nucleus). Combinations of dye concentration, incubation time and temperature must be tested to minimize the amount of dye that compartmentalized into organelles (tested by confocal microscopy). Experimental conditions used in this protocol have been optimized for 1321N1 astrocytoma, C6 Glioma and DI TNC cell lines.
2. Optimal fura-2 loading time and de-esterification time may be determined empirically depending on the cell type. Fura-2 AM, Pluronic F-127, Probenecid concentrations and incubation time stated in this protocol have been optimized for 1321N1 astrocytoma, C6 Glioma and DI TNC cell lines.
3. Receptor agonist solution preparation:
  - a. Agonist solution needs to be prepared taking in consideration well volume and the volume to be dispensed by the instrument injectors.
  - b. *E.g.*, Well initial volume = 200  $\mu$ l  
Volume to be dispense = 20  $\mu$ l  
Well final volume = 220  $\mu$ l  
Desired agonist concentration = 100  $\mu$ M ATP  
Agonist stock concentration = 1.1 mM ATP
4. Tecan i-Control parameters are optimized for this protocol and must be determined empirically.
5. HBS can be stored at 4 °C. Allow to reach room temperature before use.
6. Store Pluronic F-127 20% solution at room temperature. If precipitation is observed, the product may be resolubilized by heating at ~40 °C and vortexing before use.

## **Recipes**

1. HEPES buffered saline (HBS), pH 7.4
  - 145 mM NaCl
  - 5 mM KCl
  - 1 mM CaCl<sub>2</sub>
  - 1 mM MgCl<sub>2</sub>

- 10 mM HEPES
- 10 mM glucose
- 2. Dye solution
  - HEPES buffered saline (HBS), pH 7.4
  - 0.1% BSA (w/v)
  - 5  $\mu$ M fura-2 AM
  - 0.05% Pluronic F-127
- 3. Medium for wild type (WT) human 1321N1 astrocytoma cells
  - DMEM
  - 10% fetal bovine serum (FBS)
  - 1% antibiotic/antimycotic
- 4. Medium for hHA-P2Y<sub>2</sub>R expressing 1321N1 cells
  - DMEM
  - 10% fetal bovine serum (FBS)
  - 1% antibiotic/antimycotic
  - 500  $\mu$ g/ml Geneticin, G418
- 5. Low serum medium
  - DMEM
  - 1% fetal bovine serum (FBS)

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### **References**

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