

## Endocytosis Detection in *Magnaporthe oryzae*

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**[Abstract]** Endocytosis is an intracellular trafficking pathway that occurs in nutrient uptake, signal transduction and reconstruction of cell polarity and is conserved in eukaryotic cells. In fungi, endocytosis plays crucial roles in the physiology of hyphal growth and pathogenicity. Evidence for endocytosis in filamentous fungi is detected by the membrane-selective dyes FM4-64. Cells of a range of filamentous fungal species readily take up these dyes. However, the method for endocytosis detection has not been well established in *Magnaporthe oryzae*. Here, we provide a protocol for tracking endocytosis in *Magnaporthe oryzae*.

**Keywords:** Endocytosis, Intracellular trafficking, Fungi, FM4-64, Hyphal growth, Pathogenicity, *Magnaporthe oryzae*

**[Background]** Endocytosis is an important cellular process that internalizes extracellular materials and retrieves membrane and proteins from the plasma membrane. Endocytosis starts from the plasma membrane and transports proteins, lipids and molecules from the external environment to degradative organelles, or recycled back to the plasma membrane by vesicles and endosomes (Kaksonen *et al.*, 2003). Several studies have demonstrated the existence of endocytosis with the fluorescent dye FM4-64, a specific tracer of endocytosis, suggesting this organelle may be involved in endocytic membrane recycling in filamentous fungi (Fischer-Parton *et al.*, 2000; Wedlich-Soldner *et al.*, 2000; Penalva, 2005; Li *et al.*, 2017). FM4-64 is also used for detecting membrane traffic from endosomes toward the vacuole and autophagy in yeast (Hayden *et al.*, 2013; Journo *et al.*, 2008). Many protocols for detecting endocytosis have been established in *Aspergillus oryzae* and yeast (Higuchi *et al.*, 2011; Higuchi *et al.*, 2009; Kamble *et al.*, 2011). However, there is no specific protocol established for *Magnaporthe oryzae*, which is the causal agent of rice blast, the most serious fungal disease in the world. Here, we describe a reliable and simple protocol for detecting the endocytic pathway of *M. oryzae*.

### **Materials and Reagents**

1. Microscope slide (Sail brand, catalog number: 7105)
2. Microscope cover glass (Fisher brand, catalog number: 12-540-A)
3. Hyphae of *Magnaporthe oryzae*

4. *M. oryzae* Guy11, used as the parental wild type strain. (*MoARK1* gene knock-out mutant  $\Delta Moark1$  was obtained in our previous work. All the strains were cultured on complete medium [CM] agar plates. Liquid CM medium was used to prepare the mycelia for endocytosis assay. The strain Guy11 was stored in our laboratory.)
5. N-(3-triethylammoniumpropyl)-4-(pdiethyl-aminophenyl-hexatrienyl) pyridinium dibromide FM4-64 (Invitrogen, catalog number: T13320)
6. Latrunculin B (Cayman, catalog number: 10010631)
7. Double-distilled H<sub>2</sub>O (ddH<sub>2</sub>O)
8. DMSO (Sigma, catalog number: D2650)
9. D-glucose (Sigma, catalog number: 47829)
10. Peptone (Sangon biotech, catalog number: A505247)
11. Yeast extract (Oxoid, catalog number: LP0021)
12. Casamino acid (Sangon biotech, catalog number: A603060)
13. Biotin (Sigma, catalog number: B4501)
14. Pyridoxin (Sigma, catalog number: P9755)
15. Thiamine (Sangon biotech, catalog number: A600939)
16. Riboflavin (Sangon biotech, catalog number: A600470)
17. p-aminobenzoic acid (Amresco, catalog number: 0779)
18. Nicotinic Acid (Sigma, catalog number: 72309)
19. Liquid complete medium (Liquid CM medium) (see Recipes)
20. Vitamin solution (see Recipes)

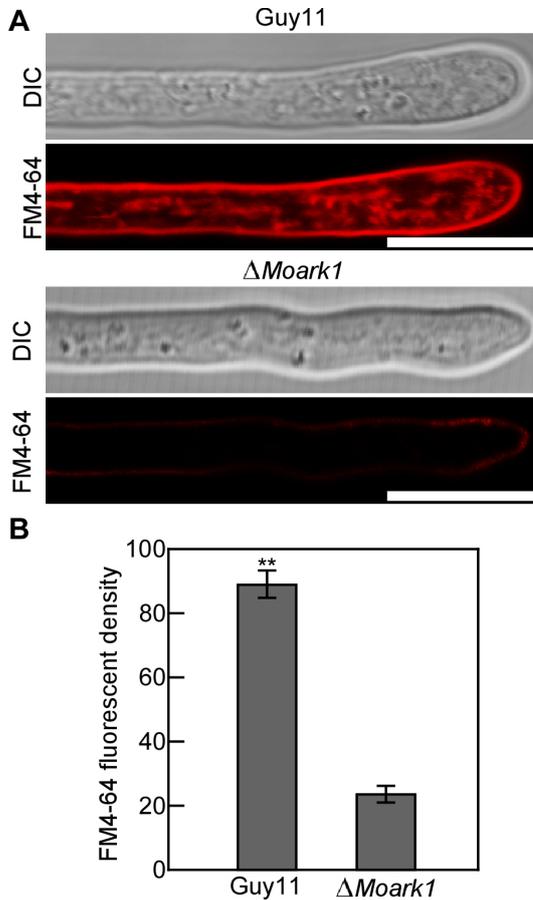
## **Equipment**

1. Eppendorf micropipette (1,000  $\mu$ l, 100  $\mu$ l, 10  $\mu$ l)
2. Confocal fluorescence microscope (Zeiss LSM710, 63x oil)

## **Procedure**

1. Culture the wild type strain Guy11 and  $\Delta Moark1$  on solid CM medium for 4 days at 28 °C. Cut the agar culture into 1 mm x 1 mm squares and culture the squares in liquid CM for 2 more days.
2. Prepare a stock solution of FM4-64 (1.3 mg/ml in DMSO) and dilute in distilled H<sub>2</sub>O at a final concentration of 5  $\mu$ g/ml, keep this staining solution on ice.
3. Wash the hyphae with distilled water and stain with FM4-64 on glass slide, about 20  $\mu$ l working solution dropped on the hyphae and stain for 2 min at room temperature.
4. Before observation, wash the hyphae with ddH<sub>2</sub>O.
5. Take photographs under a confocal fluorescence microscope (63x oil, 570/620 nm excitation, 590 nm dichroic, 630/660 nm emission is used to observe the fluorescence of FM4-64) after

exposure to FM4-64. A representative picture of the plasma membrane and the inner cellular membrane components after staining 2 min with FM4-64 is shown in Figure 1.



**Figure. 1 Endocytosis assay in *M. oryzae*.** A. The wild type strain Guy11 and the  $\Delta Moark1$  were grown in liquid CM for 48 h before the addition of FM4-64 and photographs were taken under a confocal fluorescence microscope (Zeiss LSM710, 63x oil) after 2 min of exposure to FM4-64. Bars = 10  $\mu$ m. B. The calculation of relative fluorescent density. Error bars represent SD and double asterisks indicate statistically significant differences (\*\*,  $P < 0.01$ ).

6. Prepare a stock solution of Latrunculin B (Lat B) in DMSO at a concentration of 25 mg/ml and dilute in distilled H<sub>2</sub>O at a final concentration of 0.1  $\mu$ g/ml. Lat B is used as an actin polymerization inhibitor to disrupt endocytosis (Kaksonen *et al.*, 2003).
7. For Lat B staining, pretreat the hyphae with Lat B when cultured in Step 1 for 30 min prior to the addition of FM4-64.
8. Repeated Steps 2-5.

### Data analysis

For endocytosis defect analysis, culture the strains Guy11 and the gene knock-out mutants in the

background of Guy11 for 2 days on microscope slides overlaid with complete medium (CM) before staining with FM4-64 dye and take photographs at various times after FM4-64 exposure, and the signal appeared on the plasma membrane and endomembrane compartments in Guy11, but did not occur or delayed until 20-30 min after staining in the mutant cells (Li *et al.*, 2017). Each result is presented at least three replicated measurements. The significance of differences between treatments is statistically evaluated using SDs and one-way analysis of variance (ANOVA) in SPSS 2.0 (<https://spss.en.softonic.com/>, Chicago, IL, USA). Data for two specific different treatments are compared statistically using ANOVA, followed by an F-test if the ANOVA result is significant at  $P < 0.05$  or  $P < 0.01$ .

### **Recipes**

1. Liquid CM medium

10 g D-glucose

2 g peptone

1 g yeast extract

1 g casamino acid

1 ml Vitamin Solution

Hygrothermal high pressure sterilization at 121 °C for 20 min.

2. Vitamin Solution

0.01 g Biotin

0.01 g Pyridoxin

0.01 g Thiamine

0.01 g Riboflavin

0.01 g p-aminobenzoic acid

0.01 g Nicotinic Acid

Add ddH<sub>2</sub>O to 100 ml and store in a dark glass bottle at 4 °C

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### **Competing interests**

No conflicts of interest or competing interests.

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