

## Intracellular cAMP Measurements in *Candida albicans* Biofilms

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**[Abstract]** *Candida albicans* is the most common cause of fungal infections worldwide. Infection by *C. albicans* is closely associated with its ability to form a biofilm, closely packed communities of cells attached to the surfaces of human tissues and implanted devices, in or on the host. When tested for susceptibility to antifungals, such as polyenes, azoles, and allylamines, *C. albicans* cells in a biofilm are more resistant to antifungal agents than *C. albicans* cells in the planktonic form. Cyclic Adenosine monophosphate (cAMP) is one of the key elements for triggering hyphal and biofilm formation in *C. albicans*. It is hard to detect or extract molecular markers (e.g., cAMP) from *C. albicans* biofilms because the biofilms have a complex three-dimensional architecture with an extracellular matrix surrounding the cell walls of the cells in the biofilm. Here, we present an improved protocol that can effectively measure the level of intracellular cAMP in *C. albicans* biofilms.

**Keywords:** *Candida albicans*, Biofilm, Second messenger, Cyclic Adenosine monophosphate, cAMP enzyme immunoassay, Enzyme linked immunosorbent assay

**[Background]** *Candida albicans* is a member of the fungal community in the human gastrointestinal tract, mouth, and genital tract. *C. albicans* does not only cause superficial diseases, but also systemic infections in immunocompromised patients (Brown *et al.*, 2012; Kim, 2016; Lohse *et al.*, 2018). In clinical infections of *C. albicans*, the fungal cells commonly form biofilms; *C. albicans* biofilms are associated with chronic infection and antifungal resistance (Rajendran *et al.*, 2016). The cyclic adenosine monophosphate (cAMP) signaling pathway has been considered to be one of the most important signal transduction pathways that can cause a switch from budding to hyphal morphogenesis, a switch to hyphal growth promotes biofilm formation and invasive growth in *C. albicans* (Wang, 2013; Tao *et al.*, 2017). Activation of adenylyl cyclase to produce cAMP is thought to promote hyphal induction, and maximal levels of cAMP coincide with maximal germ tube formation (Parrino *et al.*, 2017). In brief, it is generally accepted that biofilm formation of *C. albicans* requires elevated levels of cAMP. Therefore, measurement of cAMP levels in *C. albicans* biofilms is important to understand biofilm formation and antifungal drug resistance and has the potential to be an important reference value for future drug development efforts.

*C. albicans* biofilms are comprised of complex, surface-associated cell populations embedded in an extracellular matrix (Lohse *et al.*, 2018). The three-dimensional hyphal structure of biofilms and the matrix surrounding the cells serve as physical barriers that make it difficult to detect or extract specific molecular markers such as cAMP from *C. albicans* biofilms. Yun *et al.* (1998) presented a method of to

measure the intracellular cAMP level of planktonic *Saccharomyces cerevisiae*; however, no protocol has been reported on how to measure the level of intracellular cAMP in *C. albicans* biofilms. This article provides an improved protocol to measure the intracellular cAMP level of *C. albicans* biofilms. In this protocol, the levels of intracellular cAMP of cells within biofilms are examined by using an enzyme-linked immunosorbent assay (ELISA) with modifications; this is a modification of the method originally described in Yun *et al.* (1998).

## **Materials and Reagents**

1. Stainless steel tweezer (Thermo Fisher Scientific, Thermo Scientific™, catalog number: DS0399-0001)
2. Pipettes (Eppendorf, P1000, catalog number: 3120000062; P200, catalog number: 3120000054; P10, catalog number: 3120000020; P2.5, catalog number: 3120000011, respectively)
3. Flasks (Duran, 50 ml, catalog numbers: 21-216-17; 100 ml, catalog number: 21-216-24, respectively)
4. Cell culture dish (Thermo Fisher Scientific, Thermo Scientific™, Nunc™ EasYDish™, catalog number: 150466)
5. 1.5 ml microtubes (Axygen®, catalog number: MCT-150-C)
6. Tubes (Eppendorf, 15 ml, catalog number: 0030122151; 50 ml, catalog number: 0030122178, respectively)
7. Cell scraper (Costar, catalog number: 3010)
8. Pipette tips (Eppendorf, 1,000 µl, catalog number: 0030000919; 200 µl, catalog number: 0030000870; 10 µl, catalog number: 0030000811, respectively)
9. Aluminum foil
10. Parafilm
11. Strain (*Candida albicans*, SC5314) (ATCC, catalog number: 10231)
12. Yeast extract (BD Biosciences, catalog number: 212750)
13. Peptone (BD Biosciences, catalog number: 211677)
14. Dextrose (BD Biosciences, catalog number: 215530)
15. Agar (BD Biosciences, catalog number: 214040)
16. RPMI 1640 medium (Sigma-Aldrich, catalog number: R6504)
17. Sodium phosphate monobasic monohydrate (Thermo Fisher Scientific, catalog number: S369-500)
18. Sodium phosphate dibasic anhydrous (Thermo Fisher Scientific, catalog number: BP332-500)
19. Sodium chloride (Thermo Fisher Scientific, catalog number: S671-3)
20. Sodium hydroxide (NaOH) (Sigma-Aldrich, catalog number: 59223C)
21. Distilled water
22. 2-(N-Morpholino)ethanesulfonic acid (MES) (Sigma-Aldrich, catalog number: M5287)
23. Trichloroacetic acid (Sigma-Aldrich, catalog number: T9159)

24. Ice
25. Liquid nitrogen
26. YPD medium
27. Ether (Sinopharm Chemical Reagent Co., Ltd, catalog number: 10009318)
28. cAMP Enzyme Immunoassay Kit (Sigma-Aldrich, catalog number: CA200)
  - a. Goat Anti-Rabbit IgG Coated 96-well
  - b. cAMP-Alkaline Phosphatase Conjugate
  - c. cAMP EIA Antibody Rabbit Anti-cAMP
  - d. 0.1 M Hydrochloric Acid
  - e. 0.1 M HCl in water
  - f. Neutralizing Reagent
  - g. Wash Buffer Concentrate
  - h. Cyclic AMP Standard
  - i. *p*-Nitrophenyl Phosphate Substrate Solution
  - j. Stop Solution
  - k. Triethylamine
  - l. Acetic Anhydride
  - m. Plate Sealer
29. MES Buffer (see Recipes)
30. Sabouraud dextrose agar medium (SDA) (see Recipes)
31. YPD broth (see Recipes)
32. YPD agar medium (see Recipes)
33. 1x PBS (see Recipes)

## **Equipment**

1. Ultrasonic cell crusher (Celton, model number: UH-500B)
2. 30 °C shaking incubator (Infors, model number: Multitron Standard)
3. CO<sub>2</sub> incubator (ESCO, CelMate<sup>®</sup>, model number: CLM-170B-8-NF)
4. Multiwell plate shaker (Thermo Fisher Scientific, Thermo Scientific<sup>™</sup>, Sorvall<sup>™</sup> ST 40R, catalog number: 75004524)
5. Multi-mode Microplate Reader (Molecular Devices, SpectraMax M5)
6. Autoclave
7. Centrifuge

## **Procedure**

- A. Growth of *Candida albicans* and biofilm formation
  1. Subculture *Candida albicans* on Sabouraud dextrose agar medium at 35 °C.

2. Pick a single colony and inoculate 10 ml of YPD medium at 30 °C in a shaking incubator (200 rpm). Grow overnight.
3. Centrifuge the culture (2,100 x *g*, 10 min) and discard the supernatant. Use 10 ml of 1x PBS to wash the cells, centrifuge the cells for 5 min at 2,100 x *g* and discard the supernatant, repeat twice. Then, discard the supernatant and harvest the cells.
4. Re-suspend the cells and dilute them to an optical density at 600 nm (OD<sub>600nm</sub>) of 0.4 in RPMI 1640 medium (10 ml RPMI 1640 medium for per cell culture dish).
5. Inoculate the suspensions into the cell culture dishes at 37 °C without shaking (inoculum should spread to cover the entire bottom of the dish). After 2 h, gently remove the inoculum (avoid scratching away the newly forming biofilms), wash with 5 ml 1x PBS twice to remove non-adherent cells. Add 10 ml fresh RPMI 1640 medium to each dish. Then, incubate at 37 °C without shaking to form biofilms.

#### B. Process biofilm cell samples

1. Remove the medium from the dishes and gently wash the biofilms with 2 ml distilled water (slowly add distilled water using a pipet to avoid disturbing the biofilms). Add 1 ml distilled water to the dishes and scrape the dishes to isolate the biofilms, and add the samples to the 15 ml tubes. Then repeat this process, add 1 ml distilled water to collect remaining samples.
2. Centrifuge the samples for 5 min at 2,100 x *g* at room temperature and discard the supernatant. Add 1 ml MES buffer and transfer the samples to the 1.5 ml microtubes, centrifuge the samples for 5 min at 2,100 x *g*, discard the supernatant.
3. Then, dry the samples in a desiccator, add 10% trichloroacetic acid (not more than half volume of MES buffer), here we add 0.5 ml.
4. Freeze the samples in liquid nitrogen, then thaw the samples on ice, repeat this process twice. Use an ultrasonic cell crusher to sonicate the samples under chilled conditions (twice at 130 W for 2 min).
5. Add 1 ml water-saturated ether, centrifuge the samples for 5 min at 600 x *g* at room temperature and discard the supernatant. Repeat four times to make sure the trichloroacetic acid is extracted.

#### C. Measure the intracellular cAMP content (Table 1)

*Note: The cAMP content was measured with a cAMP enzyme immunoassay system following the manufacturer's instructions. Allow all reagents to warm to room temperature for at least 30 min before opening and don't use polystyrene tubes.*

**Table 1. The procedure for measurement of intracellular cAMP content**

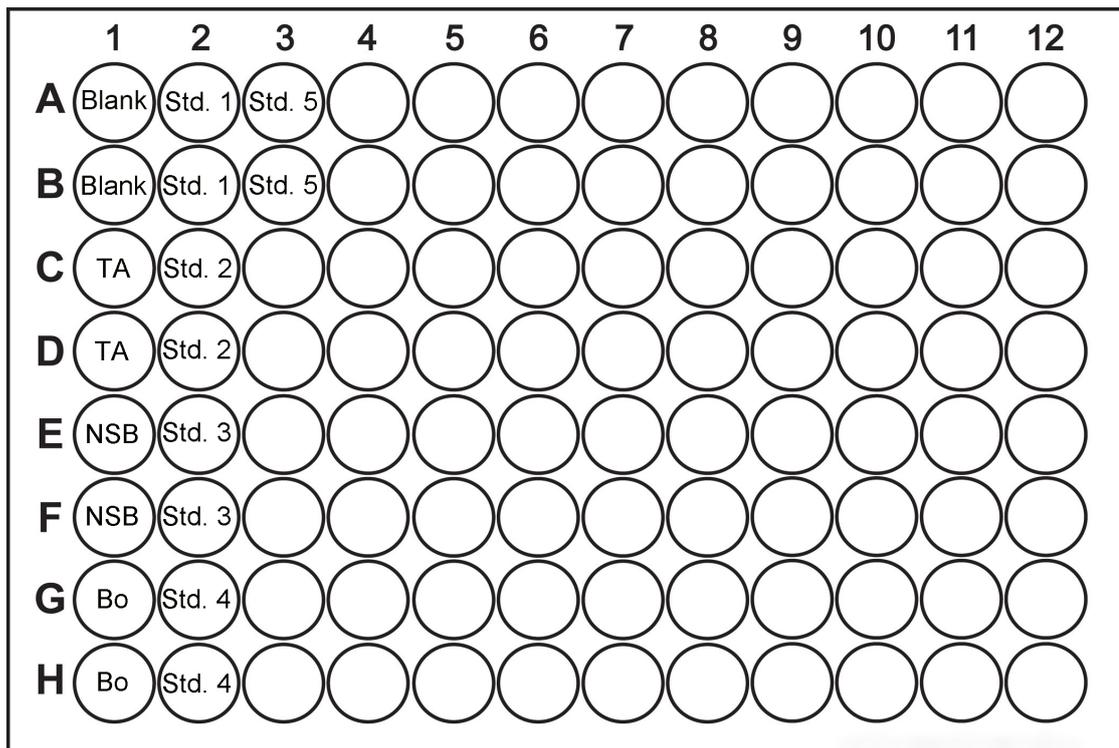
Well I.D.	Blank	TA	NSB	Zero Std. (Bo)	Std.	Samples
	A1, B1	C1, D1	E1, F1	G1, H1	A2 - B3	C3 - H12
Neutralizing Reagent	-	-	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l
0.1 M HCl	-	-	150 $\mu$ l	100 $\mu$ l	-	-
Std. and/or Sample	-	-	-	-	100 $\mu$ l	100 $\mu$ l
Conjugate	-	-	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l
Antibody	-	-	-	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l
Incub. 2 h at RT, shaking				--		
Asp. & Wash 3 x 200 $\mu$ l				--		
Conjugate	-	5 $\mu$ l	-	-	-	-
Substrate	200 $\mu$ l	200 $\mu$ l	200 $\mu$ l	200 $\mu$ l	200 $\mu$ l	200 $\mu$ l
Incub. 1 h at RT				--		
Stop Solution	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l	50 $\mu$ l

1. Add 0.5 ml of acetic anhydride to 1 ml of triethylamine to prepare the Acetylation Reagent. Dilute 10 ml Wash Buffer Concentrate with 90 ml distilled water to prepare 1x Wash Buffer.
2. Prepare cAMP Standard solutions:
  - a. Label five 12 x 75 mm tubes from 1 to 5.
  - b. Add 990  $\mu$ l of 1 M HCl into tube 1 and 750  $\mu$ l into tubes 2-5. Add 10  $\mu$ l of the Cyclic AMP Standard to tube 1 and mix thoroughly.
  - c. Then, transfer 250  $\mu$ l solution from Tube 1 to Tube 2. Dilute the cAMP from Tube 2 to Tube 5 in series. The concentration of cAMP in Tubes 1 through 5 will be 20, 5, 1.25, 0.312, and 0.078 pmol/ml, respectively (Table 2).
  - d. Label one 12 x 75 mm tube as the Zero Standard/NSB tube.

**Table 2. Preparation of cAMP standard solution**

Standard	0.1 M HCl Added ( $\mu$ l)	cAMP Standard Added ( $\mu$ l)	cAMP (pmol/ml)
1	990	10, Stock	20
2	750	250, Std. 1	5
3	750	250, Std. 2	1.25
4	750	250, Std. 3	0.312
5	750	250, Std. 4	0.078

3. Add 50  $\mu$ l Neutralizing Reagent into each well of the ELISA plate, except wells A1-D1 (Figure 1). Add 100  $\mu$ l 0.1 M HCl into the wells E1-H1, respectively. Add 100  $\mu$ l of standard solutions 1-5 into wells A2-B3.
4. Add 100  $\mu$ l of the samples into the appropriate wells.
5. Add 50  $\mu$ l 0.1 M HCl into wells E1-F1 (NSB wells, non specific binding wells).
6. Add 50  $\mu$ l blue cAMP-Alkaline Phosphatase Conjugate into each well except wells A1-D1. Add 50  $\mu$ l yellow cAMP EIA antibody into all of the wells except A1-F1 (Figure 1).



**Figure 1. Plate layout for the cAMP measurement**

7. Incubate the plate at RT for 2 h on a plate shaker (500 rpm).
8. Empty the contents of the wells and use 1x Wash Buffer to wash every well. Repeat twice and discard the remaining wash buffer.
9. Add 50  $\mu$ l of blue cAMP-Alkaline Phosphatase Conjugate to the TA wells. Add 200  $\mu$ l of the *p*-Nitrophenyl Phosphate Substrate Solution to every well.
10. Incubate the plate at RT for 1 h without shaking.
11. Add 50  $\mu$ l of Stop Solution to every well. Read the optical density (OD) at 405 nm, preferably with correction between 570 and 590 nm.

### **Data analysis**

Calculate the concentration of cAMP as follows (Table 3):

- a. Calculate the Average Net Optical Density (OD) bound for each standard and sample:  
Average Net OD = Average Bound OD – Average NSB OD
- b. Calculate the binding of each pair of standard wells as a percentage of the maximum binding wells (Bo):  
Percent Bound = Net OD/Net Bo OD x 100
- c. Use the Logit-Log paper plot Percent Bound versus Concentration of cAMP for the standards and draw a straight line through those points. Then, determine the concentration of cAMP by interpolation.

**Table 3. Typical results for calculating the concentration of cAMP**

Sample	Net OD	Percent Bound (B/Bo)	cAMP (pmol/ml)
Blank OD	(0.072)	-	-
TA	1.691	-	-
NSB	0.000	0%	-
Bo	0.257	100%	0
S1	0.029	11.31%	20
S2	0.064	24.95%	5
S3	0.129	50.29%	1.25
S4	0.206	80.12%	0.312
S5	0.252	98.05%	0.078

Repeat the test in triplicate for each assay, and test each group in triplicate on different days.

### **Recipes**

1. MES Buffer (1 L)  
21.3 g MES  
5 M NaOH (adjust pH to 6.0)  
1 L Distilled water  
Autoclave (120 °C, 120 kPa for 20 min)
2. SDA agar medium (1 L)  
10 g Peptone  
40 g Dextrose  
20 g Agar (2%)  
1 L Distilled water  
Autoclave (120 °C, 120 kPa for 20 min)
3. YPD broth (1 L)  
10 g Yeast extract (1%)  
20 g Peptone (2%)  
20 g Dextrose (2%)  
1 L Distilled water  
Autoclave (120 °C, 120 kPa for 20 min)
4. YPD agar medium (1 L)  
10 g Yeast extract (1%)  
20 g Peptone (2%)  
20 g Dextrose (2%)

- 20 g Agar (2%)
- 1 L Distilled water
- Autoclave (120 °C, 120 kPa for 20 min)
- 5. 1x PBS
- 2.62 g Sodium phosphate monobasic monohydrate
- 11.5 g Sodium phosphate dibasic anhydrous
- 43.84 g Sodium chloride
- 500 ml Distilled water
- Adjust pH to 7.2 (1 M NaOH)
- Autoclave (120 °C, 120 kPa for 20 min)

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### **Competing interests**

The authors declare that there are no conflicts of interest or competing interests.

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