

## Isolation of Healthy F4/80+ Macrophages from Embryonic day E13.5 Mouse Fetal Liver Using Magnetic Nanoparticles for Single Cell Sequencing

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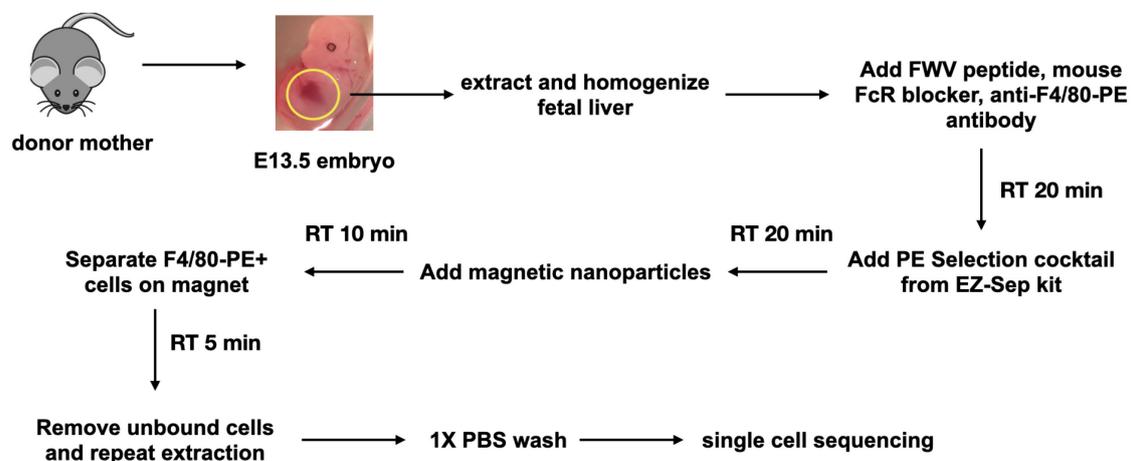
**[Abstract]** *In vivo* erythropoiesis occurs in the erythroblast island niche (EBI), comprising of a central macrophage that attaches to and aids the maturation of erythroid progenitors into mature reticulocytes. Macrophages in hematopoietic tissue such as embryonic fetal liver are heterogeneous and express the cell surface protein F4/80. Earlier methods of isolating F4/80+ macrophages from hematopoietic tissue relied on FACS sorting, but the relatively low numbers of F4/80+ cells obtained after FACS sometimes led to poor RNA quality. Additionally, since EBI macrophages are attached to erythroblasts, care must be taken to avoid contamination with bound erythroblasts. We have developed a novel method for isolating F4/80+ cells from E13.5 mouse fetal liver using magnetic nanoparticles, which can be performed on the lab bench. During cell suspension and homogenization, we also add a peptide that disrupts erythroid macrophage interactions and generates F4/80+ single cells free of erythroid contamination. Thus, our protocol generates a population enriched in F4/80+ cells that are healthy and ready for sensitive techniques such as single cell sequencing.

**Keywords:** Erythroblast island macrophages, F4/80, EZ-sep magnetic nanoparticles, Single cell sequencing, Fetal liver

**[Background]** Single cell sequencing technology has revolutionized our understanding of cell populations in a manner where cell types earlier perceived as homogeneous have been found to contain a great degree of heterogeneity. One such population comprises macrophages that support erythropoiesis in the erythroblast island (EBI) niche (Manwani and Bieker, 2008; Klei *et al.*, 2017; Li *et al.*, 2021). Previous studies have indicated that only a subpopulation of macrophages in erythroid tissues forms the EBI niche (Xue *et al.*, 2014; Seu *et al.*, 2017; Li *et al.*, 2019), and single cell sequencing would be an excellent way to address the question of macrophage heterogeneity in erythropoietic tissue. However, since the EBI macrophage forms tight adhesive interactions with the maturing erythroblasts, removing erythroblasts from isolated macrophage populations is critical to the obtention of a pure population of macrophages for analysis.

Earlier efforts to isolate macrophages from hematopoietic tissues relied on using the macrophage-

specific cell surface protein F4/80 as a marker for FACS-based methods (Xue *et al.*, 2014; Li *et al.*, 2019). However, macrophages comprise a relatively smaller fraction of the total cells in E13.5 fetal liver, and our previous experience with FACS sorting these cells showed that they resulted in poor quality of RNA or unhealthy cells that did not grow well in culture. Thus, for single cell sequencing, we decided to apply an isolation protocol using magnetic nanoparticles that usually generates healthier cells without compromising on purity (Figure 1). To reduce erythroid contamination in the final isolated F4/80+ macrophage population, during cell homogenization, we used a peptide that specifically inhibits the ICAM4/ $\alpha$ v interactions that form the core of erythroblast-macrophage adhesive junctions (Xue *et al.*, 2014). The EZ-Sep magnetic nanoparticle isolation kit from Cell Signaling Technologies utilizes a selection reagent that binds to phycoerythrin (PE) conjugated to a primary antibody such as anti-F4/80. The next step involves selection of F4/80-PE+ cells using the EZ-Sep PE selection cocktail (EZ-Sep mouse PE selection kit from Cell Signaling Technologies) and subsequent isolation using magnetic nanoparticles (Figure 1); this latter step can be repeated multiple times to increase purity. However, there is a trade-off where increased repetitions result in increased cell loss, which becomes a significant factor when dealing with fewer cell numbers. Thus, we repeated the purification step twice and were able to generate approximately 85% purity as assayed by flow cytometry for F4/80; in addition, the F4/80- cells were negative for erythroid markers indicating low erythroid contamination (Mukherjee *et al.*, 2021).



**Figure 1. Schematic of F4/80-PE+ cell isolation for single cell sequencing using EZ-Sep magnetic nanoparticles.**

## **Materials and Reagents**

1. 70  $\mu$ m cell strainer (Fisher Scientific, Falcon, catalog number: 10788201)
2. Round bottom polystyrene 5 ml test tubes (Fisher Scientific, Falcon, catalog number: 14-959-5)
3. 1.7 ml microfuge tubes (Crystalgen)
4. 50 ml conical tubes (BD)

5. Sterile pipettes (Corning), tissue culture 6 cm dishes (BD)
6. Mouse E13.5 embryonic fetal liver
7. EasySep mouse PE positive selection kit (Cell Signaling Technologies, catalog number: 17656)
8. Anti-F4/80-PE antibody (rabbit polyclonal) (eBiosciences, catalog number: 12-4801-80)
9. FWV ICAM4/av inhibitor peptide (custom sequence LRQGKTLRG) (Genscript, catalog number: SC1848). A stock solution was prepared at 23 mM and stored at -20°C. Stock solution is stable for at least 2 years.
10. Cell Culture Phosphate Buffered Saline 1× (Fisher Scientific, Corning, catalog number: MT21040)
11. Fetal Bovine Serum (GE Healthcare, HyClone, catalog number: SH30070.03)
12. 0.5 M EDTA stock solution

### **Equipment**

1. Pipettes (P200)
2. Dissection tools (e.g., sharp tweezers)
3. EasyEights EasySep Magnet (Cell Signaling Technologies, catalog number: 18103)
4. Refrigerated centrifuges capable of fitting 1.7 ml and 50 ml tubes
5. Flow Cytometer (Attune)

### **Procedure**

#### A. Mouse E13.5 embryonic fetal liver extraction and cell homogenization

1. Sacrifice the mouse and extract E13.5 embryos, placing them in ice-cold 1× PBS in a sterile 6 cm tissue culture dish.
2. Using sharp tweezers, carefully extract the fetal livers into 200 µl of 1× PBS with 2% FBS.
3. Using a P200, pipette repeatedly but gently to homogenize the livers and generate single cell suspensions (collagenase or other enzymatic treatments are not required).
4. Filter the cells by passing them through a 70 µm mesh filter into 5 ml of ice-cold 1× PBS + 2% FBS in a 50 ml conical tube.
5. Mix well, take a small aliquot for counting the cells, and centrifuge the remaining cells in a refrigerated centrifuge at 180 × g for 5 min at 4°C.

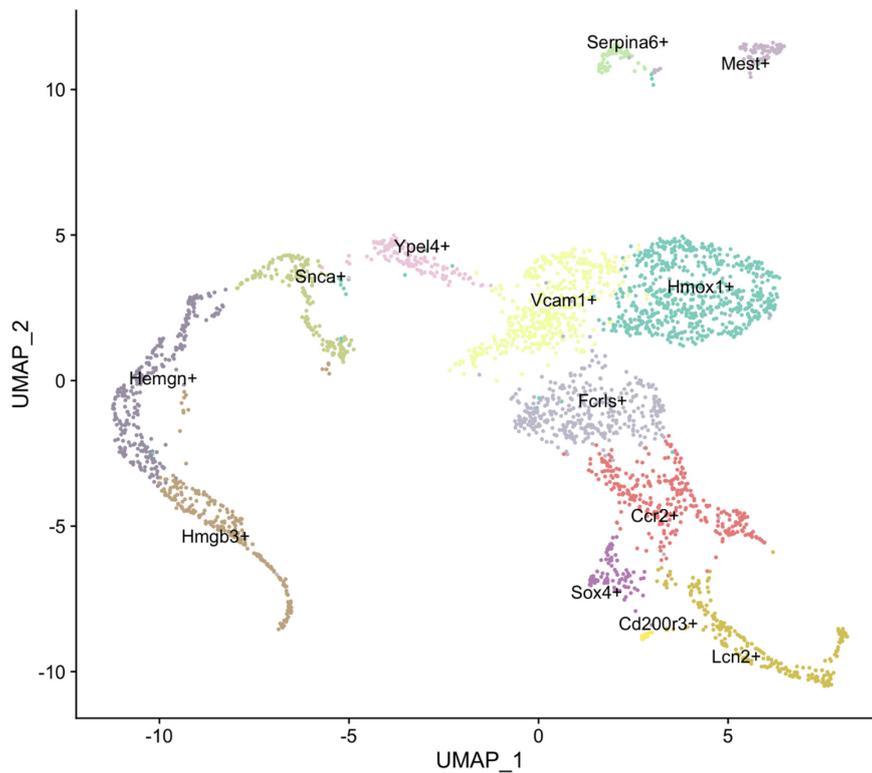
#### B. F4/80-PE+ cell selection using EZ-Sep mouse PE selection kit

1. Resuspend the cells in 1× PBS + 2% FBS + 1 mM EDTA (EZ-Sep recommended medium) at room temperature and at a density of up to 10<sup>8</sup> cells per ml in a microfuge tube. (Usually, each fetal liver generates about 5-10 million cells, so each liver can be considered one sample). We usually resuspend each fetal liver in 200 µl of 1× PBS + 2% FBS in separate tubes.

2. Add FWV peptide to a final concentration of 2 mM and mix well (~17.5  $\mu$ l of 23 mM stock solution added per 200  $\mu$ l of sample).  
*Note: At this point onwards, the steps refer to the protocol for EZ-Sep mouse PE selection kit.*
3. Add 2  $\mu$ l of mouse FcR blocker from the EZ-Sep kit and pipette to mix cells.
4. At a dilution of 1:100 (based on the total volume from Step B1), add anti-F4/80-PE antibody, mix well by pipetting gently, and incubate at room temperature in a dark place for 20 min.  
*Note: Antibody dilutions will vary depending on the antibody used and must be standardized for different antibodies.*
5. Add 2  $\mu$ l of PE selection cocktail from the EZ-Sep kit, mix well by pipetting gently, and incubate at room temperature for 20 min.
6. Add 10  $\mu$ l of magnetic nanoparticles from the EZ-Sep kit, mix well by pipetting gently, and incubate at room temperature for 10 min
7. Place the tubes on an EasyRights EasySep Magnet from Cell Signaling Technologies (which allows for eight tubes to be processed at once) for 5 min at room temperature.
8. Gently decant the supernatant by inverting the magnet with the tubes into a waste container (use one smooth inversion motion, do not shake).
9. Remove the tubes from the magnet and collect the cells in 1 ml of cold EZ-Seq recommended medium by gentle pipetting along the walls of the tubes. Centrifuge at 180  $\times$  g for 5 min at 4°C.
10. Resuspend in 200  $\mu$ l of EZ-Sep recommended medium at room temperature, and repeat the magnetic nanoparticle selection steps from Step B6.
11. Remove the tubes from the magnet and wash twice in ice-cold 1 $\times$  PBS. For each wash, add 2 ml of 1 $\times$  PBS along the walls of the tubes and place on the magnet for 5min. Decant the liquid gently each time, as in Step B8.
12. After the final wash, resuspend the cells in 0.5 ml 1 $\times$  PBS and proceed with single cell sequencing.
13. Take out a small aliquot of cells and analyze using flow cytometry for the purity of F4/80-PE+ cells.

## **Data analysis**

Please refer to Mukherjee *et al.* (2021) for details of flow cytometry data analysis of purified F4/80+ population. In this study, 28,000 healthy F4/80+ macrophages (visually counted by light microscopy after Trypan Blue staining) were isolated from two independent wild-type E13.5 fetal livers and pooled for single cell sequencing. For details of single cell sequencing quality, cell heterogeneity, and data analysis, please refer to Mukherjee *et al.* (2021). We obtained 13 independent clusters after U-MAP analysis using the Seurat package (Figure 2).



**Figure 2. U-MAP clustering of single cell RNA-Seq data of F4/80+ macrophages isolated from E13.5 mouse fetal liver.**

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This protocol was adapted with minor modification from previous study published by Mukherjee *et al.* (2021; Doi: 10.7554/eLife.61070).

### **Competing interests**

The authors declare no competing interests

### **Ethics**

This study was performed in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All animals were handled according to approved institutional animal care and use committee (IACUC) protocols (#18-1911) of

the Mount Sinai School of Medicine.

## **References**

1. Klei, T. R., Meindert, S. M., van den Berg, T. K. and van Bruggen, R. (2017). [From the Cradle to the Grave: The Role of Macrophages in Erythropoiesis and Erythrophagocytosis](#). *Front Immunol* 8: 73.
2. Li, W., Wang, Y., Zhao, H., Zhang, H., Xu, Y., Wang, S., Guo, X., Huang, Y., Zhang, S., Han, Y., *et al.* (2019). [Identification and transcriptome analysis of erythroblastic island macrophages](#). *Blood* 134(5): 480-491.
3. Li, W., Guo, R., Song, Y. and Jiang, Z. (2021). [Erythroblastic Island Macrophages Shape Normal Erythropoiesis and Drive Associated Disorders in Erythroid Hematopoietic Diseases](#). *Front Cell and Dev Biol* 8: 613885.
4. Manwani, D. and Bieker, J. J. (2008). [The erythroblastic island](#). *Curr Top Dev Biol* 82: 23-53.
5. Mukherjee, K., Xue, L., Planutis, A., Gnanapragasam, M. N., Chess, A. and Bieker, J. J. (2021). [EKLF/KLF1 expression defines a unique macrophage subset during mouse erythropoiesis](#). *Elife* 10: e61070.
6. Seu, K. G., Papoin, J., Fessler, R., Hom, J., Huang, G., Mohandas, N., Blanc, L. and Kalfa, T. A. (2017). [Unraveling Macrophage Heterogeneity in Erythroblastic Islands](#). *Front Immunol* 8: 1140.
7. Xue, L., Galdass, M., Gnanapragasam, M. N., Manwani, D. and Bieker, J. J. (2014). [Extrinsic and intrinsic control by EKLF \(KLF1\) within a specialized erythroid niche](#). *Development* 141(11): 2245-2254.